

WITHAMINIMIN, A WITHANOLIDE FROM *PHYSALIS MINIMA*

HUGO E. GOTTLIEB, MIRIAM COJOCARU, SUBHASH C. SINHA,* MAHENDRA SAHA,* ANJANA BAGCHI,* ABID ALI* and ANIL B. RAY*

Department of Chemistry, Bar-Ilan University, Ramat-Gan 52100, Israel; *Department of Medicinal Chemistry, I.M.S., Banaras Hindu University, Varanasi 221005, India

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Key Word Index—*Physalis minima*; Solanaceae; withanolides; withaminimin; steroidal lactones.

Abstract—The structure of withaminimin, a new ergostane-type steroid from *Physalis minima*, was established by spectral analysis (^1H and ^{13}C NMR, MS) and chemical transformations, as (20*S*, 22*R*)-15 α -acetoxy-5 α ,6 β ,14 α -trihydroxy-1-oxowitha-2,16,24-trienolide. An unusual MH_2O^+ quasi-molecular ion was observed in the chemical ionization mass spectrum of the natural product.

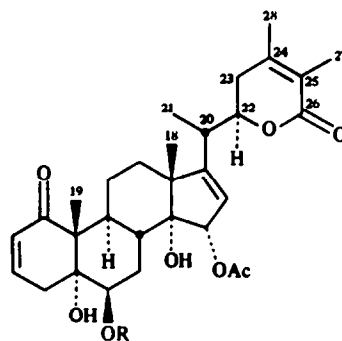
INTRODUCTION

C_{28} -Steroidal lactones based on the ergostane framework are exclusive to the Solanaceae. Of the different genera of this family that are known to yield such compounds, *Physalis* is special for elaborating complex structural variants of simpler withanolides [1, 2]. *Physalis*-derived C_{28} -steroidal lactones include withaphysalins [3], physalins [4], ixocarpalactones [5] and perulactones [6, 7], in addition to well known withanolides. While withanolides are the major constituents of *Physalis peruviana* [8, 9], no withanolide and only withaphysalins and physalins have so far been isolated from *Physalis minima* [3, 10], an annual herb growing wild in India, Sri Lanka and tropical regions of Africa. Systematic fractionation of the alcoholic extract of the plant material yielded in our hands a new withanolide for which the name withaminimin has been given. The functionalities present in the D ring of withaminimin make it a likely precursor for 13,14-secosteroids native in this plant [3, 10]. Withaminimin has also been found to be present in a poorer yield in *Physalis minima* Linn. var. *indica* C. B. Clarke [11].

RESULTS AND DISCUSSION

The ^1H NMR and ^{13}C NMR spectra of withaminimin (**1a**) (see Tables 1 and 2) were consistent with a withanolide structure, in which 1-keto- Δ^2 and 5 α ,6 β -dihydroxy functionalities were clearly present, as well as a typical α,β -unsaturated δ -lactone in the side-chain [1, 2, 12]. A negative Cotton effect at 332 nm and a positive Cotton effect at 252 nm confirmed A/B *trans*-fused rings and a 22*R*-configuration. The problem remained of locating a trisubstituted double bond, an oxymethine and an oxygenated non-protonated carbon; connected to one of these oxygens was an acetate group (carbons at δ 170.63 and 21.36, protons at 2.01). This was accomplished by selective decoupling experiments in the ^1H NMR spectrum. In the most revealing of these, irradiation at the frequency of the H-20 multiplet at δ 2.53 not only collapsed the 21-methyl signal into a singlet and the H-22 (*ddd*) into a *dd* (12 and 4 Hz), but also sharpened the doublet at δ 5.60. This suggested an allylic H-20 and therefore a 16,17 double

bond. The olefinic H-16 was in turn adjacent to the oxymethine; the strong deshielding of H-15 indicated that it belonged to a CHOAc unit. Finally, the sharp doublet for H-15 revealed the presence of a tertiary alcohol at position C-14. Inspection of models showed that the



1a R = H
1b R = Ac

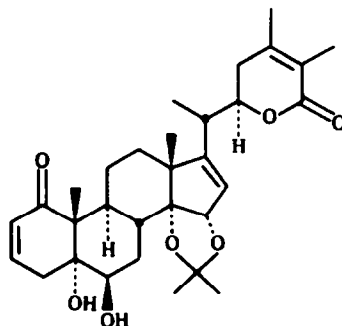


Table 1. ^1H NMR data for withaminimin (1a), withaminimin acetate (1b) and withaminimin acetonide (2)


| H | 1a | 1b | 2 |
|---|-------------------------------|-----------------------------------|-------------------------------------|
| 2 | 5.87 (<i>dd</i> , 10, 2.5) | 5.92 (<i>dd</i> , 10, 2.5) | 5.90 (<i>dd</i> , 10.5, 3) |
| 3 | 6.58 (<i>ddd</i> , 10, 5, 2) | 6.56 (<i>ddd</i> , 10, 5.5, 2.5) | 6.63 (<i>ddd</i> , 10.5, 5.5, 2.5) |
| 4 α | | | 2.10 (<i>dd</i> , 20, 5.5) |
| 4 β | 3.19 (<i>dt</i> , 20, 2.5) | | 3.34 (<i>dt</i> , 20, 3) |
| 6 | 3.60 (<i>br t</i> , 2) | 5.17 (<i>br</i> , $W_{1/2}$ = 5) | 3.78 (<i>br</i> , $W_{1/2}$ = 7) |
| 7 α | | | 1.84 (<i>td</i> , 14, 3.5) |
| 7 β | | | 2.00 (<i>dt</i> , 14, 3) |
| 8 | | | 2.30 (<i>ddd</i> , 14, 11.5, 3) |
| 9 | | | 2.13 (<i>m</i>) |
| 11–12 | | | 1.5–1.65 (<i>m</i> , 4H) |
| 15 | 5.33 (<i>d</i> , 2.5) | 5.51 (<i>d</i> , 2) | 5.15 (<i>d</i> , 2) |
| 16 | 5.60 (<i>br d</i> , 2.5) | 5.58 (<i>d</i> , 2) | 5.70 (<i>d</i> , 2) |
| 18 | 1.16 (<i>s</i> , 3H) | 1.14 (<i>s</i> , 3H) | 1.15 (<i>s</i> , 3H) |
| 19 | 1.21 (<i>s</i> , 3H) | 1.25 (<i>s</i> , 3H) | 1.40 (<i>s</i> , 3H) |
| 20 | 2.53 (<i>m</i>) | | 2.44 (<i>quintet</i> , 7) |
| 21 | 1.12 (<i>d</i> , 7, 3H) | 1.23 (<i>d</i> , 7, 3H) | 1.08 (<i>d</i> , 7, 3H) |
| 22 | 4.34 (<i>ddd</i> , 12, 7, 4) | 4.34 (<i>dt</i> , 13, 4.5) | 4.29 (<i>ddd</i> , 12.5, 6.5, 3.5) |
| 23 α | 2.31 (<i>dd</i> , 17, 4) | | 2.26 (<i>dd</i> , 17.5, 3.5) |
| 23 β | 2.42 (<i>dd</i> , 17, 12) | | 2.48 (<i>dd</i> , 17.5, 12.5) |
| 27 | 1.87 (<i>br s</i> , 3H) | 1.87 (<i>br s</i> , 3H) | 1.87 (<i>br s</i> , 3H) |
| 28 | 1.98 (<i>br s</i> , 3H) | 1.93 (<i>br s</i> , 3H) | 1.93 (<i>br s</i> , 3H) |
| 6-OAc | | 2.12 (<i>s</i> , 3H) | |
| 15-OAc | 2.01 (<i>s</i> , 3H) | 2.03 (<i>s</i> , 3H) | |
|  | | | 1.45 (<i>s</i> , 3H) |
| | | | 1.41 (<i>s</i> , 3H) |

Table 2. ^{13}C NMR data for withaminimin (1a) and its acetonide (2)

| C | 1a | 2 | C | 1a | 2 |
|----|--------|--------|-----------------------------------|--------|--------|
| 1 | 204.06 | 204.16 | 18 | 16.80 | 20.69 |
| 2 | 128.71 | 128.46 | 19 | 15.10 | 14.02 |
| 3 | 141.33 | 141.61 | 20 | 36.11 | 35.15 |
| 4 | 36.03 | 35.67 | 21 | 17.21 | 17.61 |
| 5 | 77.22 | 77.51 | 22 | 78.47 | 79.16 |
| 6 | 74.26 | 74.75 | 23 | 32.35 | 32.58 |
| 7 | 26.47 | 30.54 | 24 | 150.25 | 148.33 |
| 8 | 35.43 | 31.04 | 25 | 121.44 | 122.19 |
| 9 | 35.43 | 35.15 | 26 | 167.52 | 166.23 |
| 10 | 52.20* | 54.25† | 27 | 12.37 | 12.44 |
| 11 | 23.18 | 20.88 | 28 | 20.64 | 20.41 |
| 12 | 38.82 | 33.87 | CH_3CO | 21.36 | |
| 13 | 52.23* | 54.23† | CH_3CO | 170.63 | |
| 14 | 82.28 | 93.77 | $\text{O}-\text{C}(\text{Me})_2$ | | 27.97 |
| 15 | 83.36 | 84.46 | $\text{O}-\text{C}(\text{Me})_2$ | | 29.28 |
| 16 | 120.37 | 126.18 | $\text{O}_2\text{C}(\text{Me})_2$ | | 110.43 |
| 17 | 161.30 | 154.00 | | | |

*†Signals with the same superscript may be interchanged.

relatively small (2.5 Hz) coupling constant between hydrogens at C-15 and C-16 is consistent only with an α -acetate moiety, leaving a quasi-axial β -proton at C-15.

The ^{13}C NMR data presented in Table 2 confirmed structure 1a for withaminimin. Indeed, the carbons of rings A and B are virtually identical to those of other

withanolides with this substitution pattern [12]; the 14 α -OH group deshields C-8 (a β -effect) and shields C-7 and C-9 (γ -effects), as expected [12]. Withaminimin is, therefore, (2*S*,22*R*)-15 α -acetoxy-5 α ,6 β ,14 α -trihydroxy-1-oxo-witha-2,16,24-trienolide. The natural product can be converted to the 6-acetate 1b by treatment with acetic anhydride–pyridine (see Experimental).

In an attempt to further substantiate the proposed structure, withaminimin was treated with 8 NH_4SO_4 in acetone (see Experimental), conditions which usually result in elimination of the 14-OH group as water [8, 13]. The product, however, proved to be the acetonide, 2 (two extra methyl groups and a dioxygenated sp^3 -hybridized carbon at 110.4, see Tables 1 and 2). The formation of 2 is only consistent with a *cis*-relationship between the C-14 and C-15 oxyfunctions, and is accompanied by relatively minor conformational changes, as reflected in the NMR data shown in Tables 1 and 2.

The chemical ionization mass spectrum (CI-MS) of withaminimin (reagent gas: isobutane) showed the expected MH^+ quasi-molecular ion at m/z 529; in addition, a weak MC_3H_7^+ at m/z 571 is also seen, confirming that the molecular weight of 1a is 528. Some fragmentation of the MH^+ ion was also present (Table 3); the ion lost, in sequence, a molecule of acetic acid and three molecules of water. Two low-mass fragments were also noteworthy: m/z 125, from the side-chain lactone moiety (see e.g. refs [14, 15]), and m/z 61 (protonated acetic acid). All this refers to a mass spectral examination in which 1a was dissolved in CHCl_3 , some of the solution was transferred to a sample vial, and the solvent was dried with a flow of N_2 . Interestingly, if a crystal of the natural product was introduced directly into the sample vial, the resulting mass

Table 3. CI-MS data* for withanimin (1a) and its acetonide (2)

| Ion | 1a† | 1a‡ | 2 |
|--|--------------|--------------|--------------|
| [MH ₂ O] ⁺ = A ⁺ | | 546 (0.29) | 544 (3.12) |
| MH ⁺ = B ⁺ | 529 (2.14) | 529 (0.02) | |
| [B - H ₂ O] ⁺ | | 511 (2.37) | |
| [A - H ₂ O - Me] ⁺ | | | 511 (9.29) |
| [B - 2H ₂ O] ⁺ | | 493 (0.02) | |
| [A - AcOH] ⁺ | | 486 (0.29) | |
| [B - AcOH] ⁺ | 469 (34.11) | 469 (38.35) | |
| [A - H ₂ O - Me - CH ₂ CO] ⁺ | | | 469 (42.81) |
| [A - AcOH - H ₂ O] ⁺ | | 468 (4.91) | |
| [B - AcOH - H ₂ O] ⁺ | 451 (100.00) | 451 (100.00) | |
| [A - 2H ₂ O - Me - CH ₂ CO] ⁺ | | | 451 (100.00) |
| [B - AcOH - 2H ₂ O] ⁺ | 433 (46.41) | 433 (37.54) | |
| [A - 3H ₂ O - Me - CH ₂ CO] ⁺ | | | 433 (69.95) |
| [B - AcOH - 3H ₂ O] ⁺ | 415 (8.11) | 415 (7.39) | |
| [A - 4H ₂ O - Me - CH ₂ CO] ⁺ | | | 415 (9.09) |
| [C ₇ H ₉ O ₂] ⁺ | 125 (3.37) | 125 (5.69) | 125 (0.69) |
| AcOH ₂ ⁺ | 61 (87.15) | 61 (16.40) | |

*The data is presented as *m/z* (rel. int.).†From CHCl₃ solution (see text).

‡From crystals (see text).

spectrum was not identical to the one described above (Table 3). While the MH⁺ peak at *m/z* 529 was present, it was considerably less abundant. There was, however, a new peak at *m/z* 546, which can only be attributed to a MH₂O⁺ quasi-molecular ion. This species then lost, in sequence, a molecule of acetic acid and one of water to give new peaks at *m/z* 486 and 468, respectively. The fragmentation of MH⁺ was the same as in the first spectrum, but in addition, a new pathway seemed to be present, involving an initial loss of two molecules of water (to *m/z* 511 and 493). From the latter ion, loss of ketene (42 mass units) would branch back to the original scheme.

For acetonide 2, the chemical ionization mass spectrum (Table 3) was independent of the sample insertion technique. In both cases, MH⁺ was not observed; instead, the MH₂O⁺ peak at *m/z* 544 was present. The fragmentation is characteristic for acetonides [16], for which molecular ions are usually absent, but [M - Me]⁺ (at *m/z* 511) and (M - Me - CH₂O)⁺ (at *m/z* 469) ions dominate. The latter species seem to lose, in sequence, three water molecules.

We are not aware of any precedents in the withanolide field for the presence of a MH₂O⁺ quasi-molecular ion; the explanation might be that most such spectra have been run in the EI mode. Two sources come to mind for this added water moiety: fragmentation from another molecule of steroid, or water of crystallization. The low probability of a bimolecular reaction under chemical ionization conditions and the subtle dependence on the handling of the samples seems to favour the latter possibility.

EXPERIMENTAL

Melting points were taken with a Toshniwal apparatus and are uncorr. IR spectra were recorded on a Perkin-Elmer Infracord 137 spectrophotometer. CD was determined on a Jasco J-40 instrument. NMR spectra were run on Bruker WH-270

(270.0 MHz, ¹H for 1b) and AM-300 (300.1 MHz for ¹H, 75.5 MHz for ¹³C) spectrometers; all the data presented in Tables 1 and 2 refer to CDCl₃ solns. Chemical shifts are given in ppm downfield from internal TMS and coupling constants are in Hz. The MS were obtained by direct probe using a Finnigan 4020 quadrupole instrument. The operating conditions for chemical shift ionization were as follows: reagent gas: isobutane, ionization potential: 58 eV, emission current: 0.25 mA, electron multiplier: 1.8 kV, source temp.: 260°. TLC used chromatoplates coated with BDH silica gel G (Glaxo Laboratories, India) and CC used BDH silica gel (60–120 mesh) and neutral Al₂O₃.

Plant material. The aerial parts of *Physalis minima* Linn. were purchased from United Chemical & Allied Products, Calcutta, India and a specimen sample is being preserved.

Isolation. The air-dried epigeal parts (5.0 kg) were ground, defatted with petrol (bp 60–80°) and extracted with 95% EtOH in a Soxhlet apparatus. The extract was concd under red. pres. to ca 2.5 l, diluted with an equal vol. of H₂O, and extracted first with petrol and then with CHCl₃. The CHCl₃ layer was washed and dried (Na₂SO₄) to give, on removal of solvent, a greenish residue (ca 40 g). This was chromatographed over silica gel and eluted first with C₆H₆ and then with increasing amounts of EtOAc in C₆H₆. C₆H₆-EtOAc (3:1) eluates yielded a solid residue (8 g) which was rechromatographed over EtOAc-washed Al₂O₃. Petrol-EtOAc (1:1) eluates from this column afforded withanimin (1a) (0.65 g) as a homogeneous amorphous powder, mp 208°, IR ν_{max}^{CHCl₃} cm⁻¹: 3400, 1715 and 1680. CD (MeOH): Δε₃₃₂ -0.08, Δε₂₅₂ +0.22. EI-MS, *m/z* (rel. int.): 513 (0.3), 495 (0.3), 469 (1.6), 450 (5.1), 355 (4.5), 343 (3.4), 337 (1.6), 325 (14.2), 171 (14.6), 135 (21.2), 125 (100.0).

Withanimin acetate (1b). A mixture of withanimin (50 mg), Ac₂O (1 ml) and pyridine (0.5 ml) was kept under dry conditions for 12 hr. The usual work-up followed by CC yielded 1b (45 mg), IR ν_{max}^{KBr} cm⁻¹: 3600, 1725, 1715, 1680, 1260 and 1240.

Withanimin acetonide (2). A soln of withanimin (0.1 g) in dry Me₂CO (20 ml) was treated with 8 N H₂SO₄ (2 ml) and stirred for 24 hr at room temp. The solvent was removed from the reaction mixture at room temp., and the residue was diluted with

H₂O and quickly extracted with CHCl₃. The washed, dried and condensed CHCl₃ extract, on chromatography over silica gel and crystallization from Me₂CO, yielded fine needles of 2 (0.06 g), mp 179–181°. EI-MS, *m/z* (rel. int.): 511 (66.6), 493 (2.7), 469 (3.0), 451 (30.8), 433 (19.9), 415 (10.8), 397 (4.3), 327 (12.6), 291 (10.8), 263 (16.0), 185 (21.2), 171 (53.2), 153 (29.7), 135 (50.0), 125 (100.0).

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